



Hearing Research 168 (2002) 1-11

www.elsevier.com/locate/heares

Review

Serotonin in the inferior colliculus

Laura M. Hurley ^{a,*}, Ann M. Thompson ^b, George D. Pollak ^a

^a 1001 E. Third St., Jordan Hall, Indiana University, Bloomington, IN 47405, USA ^b Department of Otorhinolaryngology, University of Oklahoma, Oklahoma City, OK 73104, USA

Received 29 August 2001; accepted 8 December 2001

Abstract

It has been recognized for some time that serotonin fibers originating in raphe nuclei are present in the inferior colliculi of all mammalian species studied. More recently, serotonin has been found to modulate the responses of single inferior colliculus neurons to many types of auditory stimuli, ranging from simple tone bursts to complex species-specific vocalizations. The effects of serotonin are often quite strong, and for some neurons are also highly specific. A dramatic illustration of this is that serotonin can change the selectivity of some neurons for sounds, including species-specific vocalizations. These results are discussed in light of several theories on the function of serotonin in the IC, and of outstanding issues that remain to be addressed. © 2002 Elsevier Science B.V. All rights reserved.

Key words: 5HT, 5-hydroxytryptamine (serotonin); IC, inferior colliculus

1. Introduction

A common and useful model for viewing the workings of the inferior colliculus (IC) is as a collection of hard-wired circuits. This concept has been key in directing research in the IC. Exploration of auditory brainstem circuitry has revealed many of the numerous ascending and descending pathways to the IC and its subnuclei (for example Brugge, 1992; Irvine, 1992; Oliver and Huerta, 1992). Imposed upon this hardwired circuitry, however, are inputs from many different neuromodulatory systems originating in classically nonauditory regions of the brain. Among the panoply of neuromodulators in the IC are the indoleamine 5-hydroxytryptamine (5HT or serotonin) (Kaiser and Covey, 1997; Klepper and Herbert, 1991; Steinbusch, 1981; Thompson et al., 1994; Hurley and Thompson, 2001), the catecholamines dopamine (Paloff and Usunoff, 2000; Olazábal and Moore, 1989) and noradrenaline (Klepper and Herbert, 1991; Moore and Bloom, 1979; Wynne and Robertson, 1996), acetylcholine (Henderson and Sherriff, 1991), and peptide modulators

such as cholecystokinin (Fallon and Seroogy, 1984; Wynne et al., 1995), somatostatin (Wynne et al., 1995; Wynne and Robertson, 1997) and substance P (Nakaya et al., 1994; Wynne et al., 1995; Wynne and Robertson, 1997).

The neuromodulator in the IC which has received the most attention is serotonin (Hurley and Thompson, 2001; Kaiser and Covey, 1997; Klepper and Herbert, 1991; Steinbusch, 1981; Thompson et al., 1994). Serotonin is a neuromodulator which in other sensory and motor systems has been shown to have profound effects on neural processing, functionally reconfiguring the circuitry within these systems (i.e. Bassant et al., 1990; Eaton and Salt, 1989; Mooney et al., 1996; Rogawski and Aghajanian, 1980; Sillar et al., 1998; Waterhouse et al., 1986). In this review we bring together different sources of information on the projection patterns, sources, and effects of serotonin on the response properties of IC neurons, and discuss their potential relevance to existing theories of serotonin function.

2. Serotonin in the IC

Various histological techniques have all confirmed the presence of serotonin in the IC, and provide some

^{*} Corresponding author. Tel.: +1-812-856-1991;

Fax: +1-812-855-6705.

E-mail address: lhurley@bio.indiana.edu (L.M. Hurley).

A. Reconstructed 5HT fibers

B. 5HT fibers





C. IC neuron with 5HT varicosities



Fig. 1. Anatomy of serotonin in the IC. (A) A camera lucida reconstruction of all of the serotonergic fibers from one section of the brain of a Mexican free-tailed bat. The pattern of fiber density is similar in other mammals. (B) Photomicrograph of serotonin fibers from the dorsal cortex of the IC of bat which have been visualized immunohistochemically. (C) A close-up of serotonin-containing varicosities (indicated by arrows) immediately apposed to a counter-stained cell body in the IC. Scale bars = 100 μ m in A, 20 μ m in B, 10 μ m in C. DC = dorsal cortex, EC = external cortex, ICc = central IC, 5HT = 5-hydroxytryptamine = serotonin. This figure was adapted from Hurley and Thompson, 2001.

of the strongest evidence for its endogenous modulation of neurons. These techniques range from histofluorescence (Fuxe, 1965) and immunohistochemistry (Klepper and Herbert, 1991; Thompson et al., 1994; Hurley and Thompson, 2001; Kaiser and Covey, 1997), to in vivo measurement of serotonin using the techniques of microdialysis (Adell et al., 1991) and high-performance liquid chromatography (Cransac et al., 1998). Serotonin receptors are also present in the IC. These postsynaptic receptors have been measured by the binding of radioactive receptor-specific ligands, as well as by mRNA and immunohistochemical labeling techniques. Serotonin receptors fall into seven main families, and members of four of these, the 5HT1 (Chalmers and Watson, 1991; Pompeiano et al., 1992; Thompson et al., 1994; Wright et al., 1995), 5HT2 (Wright et al., 1995; Harlan et al., 2000), 5HT4 (Waeber et al., 1994), and 5HT7 (To et al., 1995) families, have been found in the IC.

While serotonin fibers are found in all subdivisions of the IC, they are not distributed uniformly (Fig. 1A,B; Hurley and Thompson, 2001; Kaiser and Covey, 1997; Klepper and Herbert, 1991; Thompson et al., 1994). Fibers are most dense in the dorsal cortex and external nucleus of the IC, and they are less dense in the central nucleus of the IC, especially in a ventromedial region. This basic pattern of serotonergic fibers is highly conserved among mammalian species, with similar staining patterns reported for two bat species, cat, rat, guinea pig, and bush baby (Hurley and Thompson, 2001; Kaiser and Covey, 1997; Klepper and Herbert, 1991; Thompson et al., 1994). Some serotonin receptor subtypes are also distributed non-uniformly. The serotonin 1C receptor has its greatest density in the external cortex of the IC (Wright et al., 1995), while the serotonin 1A receptor is densest in the posterior pericentral and dorsomedial subdivisions (Thompson et al., 1994), and the serotonin 7 receptor is densest in the dorsal cortex (To et al., 1995). In general, then, receptor density seems to follow the density of serotonin fibers.

Associated with serotonin fibers are periodic swellings or varicosities having the beads-on-a-string morphology typical of en passant synapses. Some of these varicosities are closely apposed to counter-stained cells at the light microscopic level, suggesting that they are associated with these cell bodies in a synaptic relationship (Fig. 1C; Hurley and Thompson, 2001), though this has not been confirmed at the electron microscopic level. In other parts of the brain, serotonin is thought to be released from these varicosities at a slight distance from postsynaptic terminals compared to classical neurotransmitter synapses, in a process known as volume release (Beaudet and Descarries, 1978; Bunin and Wightman, 1998; Vergé and Calas, 2000).

3. Origin of serotonin fibers

Most of the serotonergic neurons innervating the IC originate in the dorsal raphe nucleus, though a few are found in other raphe nuclei (Klepper and Herbert, 1991). Raphe nuclei are found along the midline of the brain from the medulla to the midbrain (Dahlstrom and Fuxe, 1964). Most, though not all, serotonergic neurons are associated with this chain of raphe nuclei; some serotonergic neurons are found more laterally (Jacobs and Azmitia, 1992). Though raphe nuclei contain many serotonergic neurons, they also contain neurons with other transmitter phenotypes, including GABA, substance P, and enkephalin (Chan-Palay, 1981; Grobecker, 1983; Bowker et al., 1983; Araneda et al., 1989; Tanaka et al., 1993). Some of these other transmitters are even found in serotonergic neurons as cotransmitters (i.e. Chan-Palay, 1981; Bowker et al., 1983; Araneda et al., 1989; Magoul et al., 1988).

Serotonergic neurons of the dorsal raphe project broadly across the forebrain, innervating many regions besides the IC. These regions include auditory and nonauditory cortices (Azmitia and Segal, 1978; Bobillier et al., 1975; Jacobs and Azmitia, 1992; Moore et al., 1978), as well as brainstem auditory nuclei like the cochlear nucleus and nuclei of the superior olivary complex (Klepper and Herbert, 1991; Thompson and Thompson, 2001; Thompson et al., 1995). Individual serotonergic neurons may even send collaterals to widely divergent regions of the brain, such as cortical and subcortical somatosensory regions (Petrov et al., 1992; Allen and Cechetto, 1994; Kirifides et al., 2001; Li et al., 2001).

Serotonergic release in the IC is potentially modulated by behavioral state and external sensory cues. In cats, the level of tonic activity of dorsal raphe neurons varies with the sleep-wake cycle, with cells firing at a higher rate during wakefulness and at a lower rate during sleep, especially during REM sleep (Trulson and Jacobs, 1979). Some dorsal raphe neurons also fire in conjunction with oral-buccal movements (Fornal et al., 1996). In addition, flashes of light and auditory stimuli can transiently activate or reset the firing of a large proportion of dorsal raphe neurons, so that sensory stimuli reliably trigger action potentials (Heym et al., 1982; Rasmussen et al., 1986; Trulson and Trulson, 1982). Anatomical connections that would support sensory responses by dorsal raphe neurons include a projection from a previously unidentified region near the cochlear nucleus and flocculus. This region is thought to be multisensory and is called the juxta-acousticofloccular fascicle (Ye and Kim, 2001).

4. Effects of serotonin in the IC

In this section we review evidence showing that exogenously applied serotonin strongly affects the responses of IC neurons to both simple tone bursts and to more complex, behaviorally relevant types of sounds. The experiments on which this section is based were performed mainly in Mexican free-tailed bats, which possess an excellent sense of hearing as well as a rich repertoire of communication and echolocation calls (Balcombe and McCracken, 1992; Gelfand and McCracken, 1986; Simmons et al., 1978, 1979). The types of complex sounds used in these studies are fre-



Fig. 2. The effects of iontophoresed serotonin on the responses of single IC neurons to tone bursts. (A) Serotonin affects the responses to tones across one neuron's entire frequency range equally, depressing the overall response dramatically. (B) Serotonin affects the responses of one neuron by depressing the responses to some frequencies entirely, but affecting the responses to other frequencies less. This figure was adapted from Hurley and Pollak, 2001.



Responses to all FM sweeps are decreased.

Fig. 3. Serotonin effects on frequency tuning and on the responses to FM sweeps are linked in neurons broadly affected by serotonin. At the top is a frequency plot for a single IC neuron illustrating that serotonin decreased the responses across the neuron's entire frequency range. Diagrams of three downward FM sweeps spanning different frequency ranges are superimposed on this plot. Below are plotted the peristimulus time histograms for the same neuron's response to all three FM sweeps in the control, serotonin, and in recovery. Serotonin suppressed the responses to all three sweeps. This figure was adapted from Hurley and Pollak, 2001.

quency-modulated (FM) sweeps and recorded speciesspecific vocalizations (Hurley and Pollak, 2001). These sorts of complex sounds are of interest because they more closely approximate sounds produced during behavior than do tone bursts. Here, we will focus primarily on responses to FM sweeps. FM sweeps are elements of the vocalizations of many animals (e.g. Kanwal et al., 1994; Bieser, 1998; Shipley et al., 1991). Downward FM sweeps have particular relevance for bats since they are similar in structure to the echolocation calls of many bats (for example Simmons et al., 1978, 1979, 1996; Kanwal et al., 1994).

One of the major effects of serotonin is to control the gain of the responses of IC neurons, which occurs in about 75% of serotonin-responsive neurons (Hurley and Pollak, 2001). In the majority of neurons, the gain control is downward, with serotonin depressing the responses of the neurons to tones across their entire fre-

quency range (Fig. 2A). For a few neurons, the gain control is positive, with serotonin increasing the responses to the same wide array of tones. In an interesting minority of about 25% of serotonin-responsive neurons, however, serotonin does not simply act as a gain control but instead alters the way the neurons filter sound. Serotonin does this by selectively targeting responses to some frequencies, but leaving responses to other frequencies relatively unaffected, as can be seen in Fig. 2B. The net result of these changes is that serotonin skews the range of frequencies to which these neurons are sensitive, changing their frequency tuning. Both of these types of serotonin effects, on gain control and frequency tuning, have consequences for the responses of neurons to FM sweeps and to species-specific communication calls. The next two figures illustrate these consequences for responses to FM sweeps.

In Fig. 3 is an example of a neuron for which serotonin had a gain control effect, decreasing the response



Fig. 4. Serotonin effects on frequency tuning and on the responses to FM sweeps are linked in neurons focally affected by serotonin. At the top is a plot of one neuron's frequency range; serotonin decreased responses at low frequencies but not at high frequencies. Superimposed on the plot are representations of three different FM sweeps spanning different frequency ranges. Below are the responses to the same FM sweeps. The response to FM sweep 1, confined to the lowest frequencies, was strongly decreased by serotonin. The response to FM sweep 2, spanning all frequencies, was slightly decreased by serotonin. The response to FM sweep 3, confined to the unaffected high frequencies, was not decreased by serotonin. This figure was adapted from Hurley and Pollak, 2001.

across its entire frequency range. The neuron in this figure also responded to three FM sweeps, differing in the range of frequencies they spanned, which can be seen superimposed on the frequency tuning plot. Serotonin decreased the responses to all of these FM sweeps, in keeping with its broad suppression of tones in the same neuron.

In Fig. 4 is an example of a neuron for which serotonin changed the frequency tuning, removing the response to lower frequencies but keeping the response to higher frequencies intact. These very focal changes in frequency tuning altered the range of FM sweeps that elicited a response from this neuron. In the control condition, this neuron also responded to three different FM sweeps. As expected, serotonin removed the response to the FM sweep that contained only the frequencies affected by serotonin (FM1). At the same time, serotonin slightly decreased the response to the FM sweep spanning both affected and unaffected frequencies (FM2), and had no effect on the response to the FM sweep containing only unaffected frequencies (FM3). When the responses to all three FM sweeps in the control and in serotonin are compared, it can be seen that serotonin changed the range of FM sweeps to which the neuron responded. That is, while the neuron responded to all three FM sweeps in the control, it responded to only two in serotonin. This close correspondence between the frequency specificity of serotonin effects and the effects of serotonin on the responses to both FM sweeps and species-specific calls is seen in most IC neurons.

Thus, for most neurons, the rules governing the relationship between serotonin effects on frequency tuning and on responses to complex sounds are straightforward. If serotonin affects the responses to tones within a certain frequency range, then it also affects the responses to FM sweeps and vocalizations that contain these frequencies, and has no effect on the responses to sounds which do not contain the affected frequencies.

However, in a small but intriguing group of neurons, there is no obvious correspondence between serotonin effects on frequency tuning and on the responses to complex sounds. An example is shown in Fig. 5. In the control, this neuron was selective for one recorded bat vocalization (Call A1) out of an array of 17, and thus did not respond to 16 other vocalizations that were played in this experiment. When serotonin was added, the neuron stopped responding to Call A1, but at the same time became selective for another vocalization (Call D2). Both vocalizations contain energy within the neuron's frequency range, as indicated by the dotted lines in the spectrograms (frequency versus time plots) in Fig. 5. The decrease in the response to Call A1 correlates with the general suppression of the response to tones which can be seen in the frequency plot at the top



Fig. 5. Serotonin effects on frequency tuning and on the responses to complex sounds are not always linked. The frequency plot at the top shows that serotonin suppressed the responses across the entire frequency range of this neuron. Below are plotted the responses to two species-specific vocalizations and the spectrograms of these vocalizations. Dashed lines indicate the frequency response range of the neuron relative to the vocalizations. In the control, the neuron responded only to the first vocalization. This figure was adapted from Hurley and Pollak, 2001.

of the figure. However, the increase in the response to Call D2 cannot be explained by referring to the frequency plot, since the frequency plot shows no increase of response at any frequency. For this neuron, as for other neurons in this class, serotonin effects do not correlate with frequency tuning and must be due to some unobserved variable.

Besides altering the magnitude of responses to sounds, serotonin also changes response latencies in some IC neurons. These sorts of changes have potential implications for a number of latency-dependent aspects of auditory processing, including binaural integration, coincidence detection, the shaping of the duration and timing of a response, and sound localization (for example Irvine et al., 1995; Ehrlich et al., 1997; Covey and Casseday, 1999; Park and Pollak, 1993; Park et al.,





Fig. 6. Serotonin changes the latencies of the responses of some IC neurons to tones and recorded vocalizations. For this single IC neuron, serotonin increased the response latency to a 20.4-kHz tone by 4 ms and increased the response latency to a recorded vocalization, Call I, by 2 ms.

1996; Galazyuk and Feng, 2001). An example is shown in Fig. 6. For this neuron, serotonin increased the latencies of responses to tone bursts and recorded vocalizations by as much as 4 ms. A change on this scale could have dramatic effects for the processing of sounds.

In summary, serotonin dramatically transforms the firing of many IC neurons in response to auditory stimuli. Serotonin affects the responses to both tone bursts and to more complex sounds, including species-specific vocalizations. These effects are in some cases a simple depression or facilitation, but in many cases are more selective and complex. Whether these effects of serotonin are correlated with the location of the recorded neurons in the different IC subdivisions is unknown, since most of these recordings were made in the central subdivision. However, the serotonin effects do not correlate well with the depth of the recording electrode in the IC, or with neural response properties such as whether the neurons are transient versus sustained or monotonic versus non-monotonic (personal observations). Serotonin effects may be more extensive than has currently been reported, since serotonin effects on many aspects of auditory processing such as binaural integration or the effects of serotonin in different subregions of the IC have not yet been investigated.

5. Specificity

Relatively small numbers of serotonergic neurons innervate widespread regions of the brain and spinal cord, and even single serotonergic neurons may project broadly (Petrov et al., 1992; Allen and Cechetto, 1994; Kirifides et al., 2001; Li et al., 2001). Even though there is some specificity in the projections of different subgroups of serotonin neurons (for example Jacobs and Azmitia, 1992; Jacobs et al., 1978; Kirifides et al., 2001), the projections of serotonin neurons are likely to be fairly diffuse (Jacobs and Azmitia, 1992). For this reason, it is commonly assumed that serotonin effects are non-specific, and that serotonin will have only one type of effect on a given population of target neurons. The actual situation is quite different. Specificity of serotonin action exists at numerous different levels in the brain, and several of these have also been found in the auditory brainstem in general and the IC in particular.

One level of specificity is in the pattern of serotonergic fibers that project to the IC. As noted in a previous section, serotonergic fibers are not uniform in density across the IC. Fibers are most dense in the external regions of the nucleus, including the dorsal and external cortices (Hurley and Thompson, 2001; Kaiser and Covey, 1997; Klepper and Herbert, 1991), and are less dense, though still plentiful, in the central nucleus. Since different IC subdivisions contain different types of neurons (Morest and Oliver, 1984; Oliver and Morest, 1984), the anatomical evidence suggests that some neuron types receive more serotonergic input and therefore may be more subject to serotonergic modulation than others.

Further levels of specificity of serotonin modulation in the IC are suggested by electrophysiological experiments. These experiments reveal different kinds of specific serotonin effects. First, only about half of IC neurons recorded respond to exogenous serotonin application, while the remainder are unaffected. Thus, there are separate populations of affected and unaffected neurons. Second, the nature of serotonin effects is different in different groups of neurons which are modulated by serotonin; serotonin decreases the response to sound in the majority of neurons, but it increases the response to sound in other neurons. Finally and most interestingly, even within the range of responses of some single neurons, serotonin selectively affects the responses to certain sounds, such as tone bursts of a given frequency range, or a subset of species-specific vocalizations (discussed above). While the cellular mechanisms of these diverse effects of serotonin are not understood, these results are intriguing, and suggest that serotonin performs specific functional tasks in different populations of IC neurons. One major caveat of this conclusion is that exogenously applied serotonin, which may not mimic endogenous patterns of serotonin release, was used in these experiments. Even so, such neuron-specific patterns of response suggest non-uniformity in the response to serotonin at some level.

Another level of specificity which has been extremely well-investigated in other regions of the brain, but hardly at all in the IC, is in the functions of different serotonin receptor subtypes and the corresponding intracellular cascades which they activate. Though four serotonin receptor types have been found in the IC, the functional consequences of having these different receptor types, and whether they mediate different effects of serotonin as measured electrophysiologically, are unclear.

Thus, although the pattern of projection of serotonin fibers to the IC may be diffuse, there are a number of anatomical and electrophysiological mechanisms which lead to specificity in the action of serotonin, both for single neurons and across the IC neuronal population.

6. Functions

A number of theories regarding serotonin function have been proposed based on the activity patterns of serotonergic neurons or the patterns of serotonin projections. Some of these theories potentially apply to or were developed in the auditory brainstem and midbrain. Here we consider whether recent electrophysiological data on the effects of exogenous serotonin on IC neurons are consistent with these theories, and what further functional consequences of serotonin are suggested by these electrophysiological data.

One of these theories addresses the role of behavioral state in the apparently opposite effects of serotonin on motor and sensory systems. Serotonin appears to strengthen or initiate the outputs in some motor systems, for example in locomotory networks (i.e. Jacobs and Fornal, 1993, 1999; Wallis, 1994; Sillar et al., 1998). Serotonin also inhibits some sensory inputs, such as nociceptive or somatosensory inputs, at a number of different levels, from the level of the brainstem and spinal cord (Jankowska et al., 1994, 1995; Lopez-Garcia, 1998) to the level of the cortex (Bassant et al., 1990; Eaton and Salt, 1989; Lopez-Garcia, 1998; Waterhouse et al., 1986). Some of the serotonergic neurons endogenously modulating these sensory and motor systems fire at higher rates during states of arousal than of non-arousal. Thus, in a state of heightened arousal, a proposed function of serotonin is to facilitate important motor outputs and to suppress non-essential sensory inputs. This has been called the motor hypothesis of serotonin function (Jacobs and Fornal, 1993). A consequence of this hypothesis is that serotonin should simply suppress the overall responsiveness of auditory neurons, including IC neurons.

In the IC, exogenously applied serotonin has been reported to depress the activity of neurons in response to sound (Faingold et al., 1991; Hurley and Pollak, 1999, 2001). As far as this goes, it supports the motor hypothesis of serotonin function. However, in some neurons, as noted above, serotonin effects are more complex than a simple gain control, since serotonin selectively depresses the responses to some sorts of sounds more than others (Hurley and Pollak, 2001). In these cases, even though serotonin depresses the responses to sounds, the end result is not a simple gain control but a change in the response range of the neurons. Moreover, in a minority of neurons, serotonin actually facilitates rather than depresses the responses to sounds. Thus, the predictions of the motor hypothesis apply for the neurons which are simply suppressed by serotonin, but not for the ones which are facilitated.

A second hypothesis regarding serotonin function in the IC grew out of patterns of serotonin staining in the auditory brainstem, and was proposed by Klepper and Herbert in 1991. The hypothesis is that serotonin strongly modulates regions which integrate inputs from auditory and non-auditory sources. Patterns of staining in the IC generally conform to this hypothesis. Serotonin fibers are denser in the dorsal and external cortices of the IC, and these regions also receive a somewhat different array of inputs than does the central IC. Like the central IC, the dorsal and external regions of the IC receive an array of inputs from lower auditory nuclei, though these inputs are reduced relative to the central nucleus (for example Oliver, 1987; Shneiderman et al., 1988; Oliver and Huerta, 1992). To a much greater degree than the central IC, the dorsal and external subdivisions of the IC also receive descending projections from the auditory cortex (for example Andersen et al., 1980; Luethke et al., 1989; Herbert et al., 1991; Winer et al., 1998). The non-central regions of the IC also receive projections from non-auditory regions, including visual regions (Itaya and Van Hoesen, 1982; Paloff et al., 1985; Hyde and Knudsen, 2000), somatosensory regions (Robards, 1979; Aitkin et al., 1981; Li and Mizuno, 1997), globus pallidus (Yasui et al., 1990; Moriizumi and Hattori, 1991; Shinonaga et al., 1992; Shammah-Lagnado et al., 1996), amygdala (Marsh et al., 1999), superior colliculus (Sato and Ohtsuka, 1996), and substantia nigra (Moriizumi et al., 1992), to name several. If the denser serotonin innervation in the noncentral regions of the IC actually translates into larger serotonin effects in these regions, then serotonin could potentially alter the integration of the auditory and non-auditory inputs in these peripheral regions of the IC. For example, serotonin could differentially gate inputs from different sources, as it seems to do in the superior colliculus (Huang et al., 1993; Mooney et al., 1996). These are issues that remain to be explored.

A third hypothesis regarding serotonin function in the IC has emerged from our electrophysiological data. The hypothesis is a consequence of serotonin's tendency to have selective effects, in modulating only some neurons and in changing the selectivity of some neurons for auditory stimuli (Hurley and Pollak, 2001). If extrapolated across the population of IC neurons, these selective effects of serotonin would result in an altered pattern of activity for a given sound. The specific spatial and temporal patterns of activity in response to a sensory stimulus may be important for encoding that stimulus across a population of neurons (for example Binz et al., 1990; Stanley et al., 1999; Covey, 2000; Doetsch, 2000; Petersen and Diamond, 2000; Friedrich and Laurent, 2001; Tsunoda et al., 2001). Thus, serotonin, in selectively altering the responses of single IC neurons to existing inputs, could alter the patterns that encode particular sounds in the IC. Similar to the motor hypothesis, the most interesting feature of such an alteration is that it would be dependent on the level of arousal and on incoming sensory cues, since the activity of serotonergic neurons innervating the IC is also dependent on these variables. The firing of serotonergic neurons by a heightened state of arousal or by a novel stimulus could therefore induce a reconfiguration of the sound-processing circuitry of the IC, causing sounds to be encoded by more specific and limited patterns of activity. Whether such changes in activity pattern are triggered by endogenous sources of serotonin (see Section 7), and what sorts of behavioral consequences they entail are issues that are not yet understood.

These hypotheses are a limited sample of the possible functions that serotonin could perform in the IC. They are not mutually exclusive; indeed, given the massive convergence of inputs in the IC, and the range of functions which have been proposed for the IC (for examples, see Brandao et al., 1994; Covey and Casseday, 1999; Li et al., 1998a,b; Braun, 2000), it is not unreasonable to imagine that serotonin may be modulating multiple processes in the IC. Generating further ideas of serotonin function, as well as confirming or refuting existing ones, must rely on future electrophysiological, anatomical, and behavioral experiments.

7. Conclusions

There has long been substantial anatomical evidence for the modulation of IC neurons by serotonin. More recently, there has been mounting electrophysiological evidence that serotonin, as well as other neuromodulators (Faingold et al., 1991; Farley et al., 1983; Habbicht and Vater, 1996), can alter auditory processing in the IC. Thus, there is evidence that serotonin is in a position to modulate IC neurons, that exogenously applied serotonin does modulate IC neurons, and that the patterns of endogenous serotonin release are likely to be linked to behaviorally important internal cues and even to external sensory cues, including auditory stimuli.

Despite these growing findings, there are still a number of major issues related to serotonin in the IC which have not been investigated at all. One of these is the effect that endogenous sources of serotonin have in the IC. There are a number of reasons that endogenous sources of serotonin might not have the same effects as exogenously applied serotonin. One is that the concentration of exogenously applied serotonin could be higher or lower than the physiological range. A second is that the release of endogenous sources of serotonin is likely to be timed to behavioral events and thus coordinated with a background of other modulatory influences onto the IC. There is some evidence that different neuromodulatory systems can influence each other, at the level of the neuromodulatory neurons themselves (i.e. Couch, 1970; Koyama and Kayama, 1993) and also at the level of target cells (i.e. Canfield and Dunlap, 1984; Funke and Eysel, 1993). Looking at serotonin effects in isolation may therefore not replicate the in vivo situation. A third outstanding issue is that endogenous serotonin release is likely to be more widespread within the IC than exogenous application, given the distribution of release sites, and release would also not be limited to the IC. Serotonergic fibers innervate most lower nuclei of the auditory brainstem which project to the IC, and serotonin has been shown to have electrophysiological effects in some of these nuclei (Ebert and Ostwald, 1992; Fitzgerald and Sanes, 1999; Wang and Robertson, 1997). Thus, serotonin would probably modulate the activity of the inputs to the IC at the same time it modulates the activity of IC neurons themselves, potentially resulting in a different pattern of effects in the IC than that observed with local application of serotonin to the IC alone.

Overall, while many interesting and perhaps unexpected aspects of serotonin in the IC have been revealed to date, many new and exciting prospects, particularly regarding the links between serotonin effects and behavior, remain to be explored.

References

Adell, A., Carceller, A., Artigas, F., 1991. Regional distribution of extracellular 5-hydroxytryptamine and 5-hydroxyindoleacetic acid in the brain of freely moving rats. J. Neurochem. 56, 709–712.

- Aitkin, L., Kenyon, C., Philpott, P., 1981. The representation of the auditory and somatosensory systems in the external nucleus of the cat inferior colliculus. J. Comp. Neurol. 196, 25–40.
- Allen, G., Cechetto, D., 1994. Serotoninergic and nonserotoninergic neurons in the medullary raphe system have axon collateral projections to autonomic and somatic cell groups in the medulla and spinal cord. J. Comp. Neurol. 350, 357–366.
- Andersen, R., Snyder, R., Merzenich, M., 1980. The topographic organization of corticocollicular projections from physiologically identified loci in the AI, AII, and anterior auditory cortical fields of the cat. J. Comp. Neurol. 191, 479–494.
- Araneda, S., Magoul, R., Calas, A., 1989. Tracing specific transmitter pathways in the rat CNS: combination of [³H]serotonin retrograde labeling with immunocytochemical detection of endogenous transmitters. J. Neurosci. Methods 30, 211–218.
- Azmitia, E., Segal, M., 1978. An autoradiographic analysis of the differential ascending projections of the dorsal and median raphe nuclei in the rat. J. Comp. Neurol. 179, 641–667.
- Balcombe, J.P., McCracken, G.F., 1992. Vocal recognition in Mexican free-tailed bats: do pups recognize mothers? Anim. Behav. 43, 79–87.
- Bassant, M.H., Ennouri, K., Lamour, Y., 1990. Effects of iontophoretically applied monoamines on somatosensory cortical neurons of unanesthetized rats. Neuroscience 39, 431–439.
- Beaudet, A., Descarries, L., 1978. The monoamine innervation of rat cerebral cortex: synaptic and nonsynaptic axon terminals. Neuroscience 3, 851–860.
- Bieser, A., 1998. Processing of twitter-call fundamental frequencies in insula and auditory cortex of squirrel monkeys. Exp. Brain Res. 122, 139–148.
- Binz, H., Zurhorst, C., Zimmermann, E., Rahmann, H., 1990. Neuronal substrates involved in processing of communicative acoustic signals in tree shrews: a 2-deoxyglucose study. Neurosci. Lett. 112, 25–30.
- Bobillier, P., Pettijean, F., Salvert, D., Ligier, M., Seguin, S., 1975. Differential projections of the nucleus raphe dorsalis and nucleus raphe centralis as revealed by autoradiography. Brain Res. 85, 205–210.
- Bowker, R., Westlund, K., Sullivan, M., Jf, W., Coulter, J., 1983. Descending serotonergic, peptidergic and cholinergic pathways from the raphe nuclei: a multiple transmitter complex. Brain Res. 288, 33–48.
- Brandao, M., Cardoso, S., Melo, L., Motta, V., Coimbra, N., 1994. Neural substrate of defensive behavior in the midbrain tectum. Neurosci. Biobehav. Rev. 18, 339–346.
- Braun, M., 2000. Inferior colliculus as candidate for pitch extraction: multiple support from statistics of bilateral spontaneous otoacoustic emissions. Hear. Res. 145, 130–140.
- Brugge, J.F., 1992. An overview of central auditory processing. In: Popper, A.N., Fay, R.R. (Eds.), The Mammalian Auditory Pathway: Neurophysiology. Springer-Verlag, New York, pp. 1–33.
- Bunin, M., Wightman, R., 1998. Quantitative evaluation of 5-hydroxytryptamine (serotonin) neuronal release and uptake: an investigation of extrasynaptic transmission. J. Neurosci. 18, 4854–4860.
- Canfield, D., Dunlap, K., 1984. Pharmacological characterization of amine receptors on embryonic chick sensory neurones. Br. J. Pharmacol. 82, 557–565.
- Chalmers, D., Watson, S., 1991. Comparative anatomical distribution of 5-HT1A receptor mRNA and 5-HT1A binding in rat brain–a combined in situ hybridisation/in vitro receptor autoradiographic study. Brain Res. 561, 51–60.
- Chan-Palay, V., 1981. Evidence for the coexistence of serotonin and substance P in single raphe cells and fiber plexuses: combined immunocytochemistry and autoradiography. Adv. Exp. Med. Biol. 133, 81–97.

- Couch, J.J., 1970. Responses of neurons in the raphe nuclei to serotonin, norepinephrine and acetylcholine and their correlation with an excitatory synaptic input. Brain Res. 19, 137–150.
- Covey, E., 2000. Neural population coding and auditory temporal pattern analysis. Phys. Behav. 69, 211–220.
- Covey, E., Casseday, J., 1999. Timing in the auditory system of the bat. Annu. Rev. Physiol. 61, 457–476.
- Cransac, H., Cottet-Emard, J., Hellstrom, S., Peyrin, L., 1998. Specific sound-induced noradrenergic and serotonergic activation in central auditory structures. Hear. Res. 118, 151–156.
- Dahlstrom, A., Fuxe, K., 1964. Localization of monoamines in the lower brain stem. Experientia 20, 398–399.
- Doetsch, G., 2000. Patterns in the brain. Neuronal population coding in the somatosensory system. Physiol. Behav. 69, 187–201.
- Eaton, S., Salt, T., 1989. Modulatory effects of serotonin on excitatory amino acid responses and sensory synaptic transmission in the ventrobasal thalamus. Neuroscience 33, 285–292.
- Ebert, U., Ostwald, J., 1992. Serotonin modulates auditory information processing in the cochlear nucleus of the rat. Neurosci. Lett. 145, 51–54.
- Ehrlich, D., Casseday, J.H., Covey, E., 1997. Neural tuning to sound duration in the inferior colliculus of the big brown bat, *Eptesicus fuscus*. J. Neurophysiol. 77, 2360–2372.
- Faingold, C.L., Gehlbach, G., Caspary, D.M., 1991. Functional pharmacology of inferior colliculus neurons. In: Altschuler, R.A. (Ed.), Neurobiology of Hearing: the Central Auditory System. Raven Press, New York, pp. 223–251.
- Fallon, J., Seroogy, K., 1984. Visual and auditory pathways contain cholecystokinin: evidence from immunofluorescence and retrograde tracing. Neurosci. Lett. 45, 81–87.
- Farley, G., Morley, B., Javel, E., Gorga, M., 1983. Single-unit responses to cholinergic agents in the rat inferior colliculus. Hear. Res. 11, 73–91.
- Fitzgerald, K., Sanes, D., 1999. Serotonergic modulation of synapses in the developing gerbil lateral superior olive. J. Neurophys. 81, 2743–2752.
- Fornal, C.A., Metzler, C.W., Marrosu, F., Ribiero-do-Valle, L.E., Jacobs, B.L., 1996. A subgroup of dorsal raphe serotonergic neurons in the cat is strongly activated during oral-buccal movements. Brain Res. 716, 123–133.
- Friedrich, R., Laurent, G., 2001. Dynamic optimization of odor representations by slow temporal patterning of mitral cell activity. Science 291, 889–894.
- Funke, K., Eysel, U., 1993. Modulatory effects of acetylcholine, serotonin and noradrenaline on the activity of cat perigeniculate neurons. Exp. Brain Res. 95, 409–420.
- Fuxe, K., 1965. Evidence for the existence of monoamine neurons in the central nervous system. IV. Distribution of monoamine terminals in the central nervous system. Acta Pysiol. Scand. 64, 39–85.
- Galazyuk, A., Feng, A., 2001. Oscillation may play a role in time domain central auditory processing. J. Neurosci. 21, RC147.
- Gelfand, D.L., McCracken, G.F., 1986. Individual variation in the isolation calls of Mexican free-tailed bat pups (*Tadarida brasiliensis mexicana*). Anim. Behav. 34, 1078–1086.
- Grobecker, H., 1983. Transmitter-peptide coexistence in the central nervous system. Eur. Neurol. 22, 38–46.
- Habbicht, H., Vater, M., 1996. A microiontophoretic study of acetylcholine effects in the inferior colliculus of horseshoe bats: implications for a modulatory role. Brain Res. 724, 169–179.
- Harlan, R., Yuan, Y., Garcia, M., 2000. Serotonin 5-HT2C receptors in central auditory pathways. ARO Abstr. 23, 113.
- Henderson, Z., Sherriff, F., 1991. Distribution of choline acetyltransferase immunoreactive axons and terminals in the rat and ferret brainstem. J. Comp. Neurol. 314, 147–163.
- Herbert, H., Aschoff, A., Ostwald, J., 1991. Topography of projec-

tions from the auditory cortex to the inferior colliculus in the rat. J. Comp. Neurol. 304, 103–122.

- Heym, J., Trulson, M., Jacobs, B., 1982. Raphe unit activity in freely moving cats: effects of phasic auditory and visual stimuli. Brain Res. 232, 29–39.
- Huang, X., Mooney, R.D., Rhoades, R.W., 1993. Effects of serotonin on retinotectal-, corticotectal-, and glutamate-induced activity in the superior colliculus of the hamster. J. Neurophys. 70, 723– 732.
- Hurley, L., Pollak, G., 1999. Serotonin differentially modulates responses to tones and frequency-modulated sweeps in the inferior colliculus. J. Neurosci. 19, 8071–8082.
- Hurley, L., Pollak, G., 2001. Serotonin effects on frequency tuning of inferior colliculus neurons. J. Neurophysiol. 85, 828–842.
- Hurley, L., Thompson, A., 2001. Serotonergic innervation of the auditory brainstem of the Mexican free-tailed bat, *Tadarida brasilien*sis. J. Comp. Neurol. 435, 77–88.
- Hyde, P., Knudsen, E., 2000. Topographic projection from the optic tectum to the auditory space map in the inferior colliculus of the barn owl. J. Comp. Neurol. 421, 143–145.
- Irvine, D., Park, V., Mattingley, J., 1995. Responses of neurons in the inferior colliculus of the rat to interaural time and intensity differences in transient stimuli: Implications for the latency hypotheses. Hear. Res. 85, 127–141.
- Irvine, D.R.F., 1992. Physiology of the auditory brainstem. In: Popper, A.N., Fay, R.R. (Eds.), The Mammalian Auditory Pathway: Neurophysiology, Vol. 2. Springer-Verlag, New York, pp. 153– 231.
- Itaya, S., Van Hoesen, G., 1982. Retinal innervation of the inferior colliculus in rat and monkey. Brain Res. 233, 45–52.
- Jacobs, B.L., Fornal, C.A., 1993. 5-HT and motor control: a hypothesis. TINS 16, 346–352.
- Jacobs, B.L., Fornal, C.A., 1999. Activity of serotonergic neurons in behaving animals. Neuropsychopharmacology 21, 9s–15s.
- Jacobs, B.L., Azmitia, E.C., 1992. Structure and function of the brain serotonin system. Physiol. Rev. 72, 165–229.
- Jacobs, B.L., Foote, S.L., Bloom, F.E., 1978. Differential projections of neurons within the dorsal raphe nucleus of the rat: a horseradish peroxidase (HRP) study. Brain Res. 147, 149–153.
- Jankowska, E., Läckberg, Z.S., Dyrehag, L.E., 1994. Effects of monoamines on transmission from Group II muscle afferents in sacral segments in the cat. Eur. J. Neurosci. 6, 1058–1061.
- Jankowska, E., Krutki, P., Läckberg, Z.S., Hammar, I., 1995. Effects of serotonin on dorsal horn dorsal spinocerebellar tract neurons. Neuroscience 67, 489–495.
- Kaiser, A., Covey, E., 1997. 5-HT innervation of the auditory pathway in birds and bats. In: Syka., J.L. (Ed.), Acoustical Signal Processing in the Central Auditory System. Plenum, New York, pp. 71–78.
- Kanwal, J.S., Matsumura, S., Ohlemiller, K., Suga, N., 1994. Analysis of acoustic elements and syntax in communication sounds emitted by mustached bats. J. Acoust. Soc. Am. 96, 1229–1254.
- Kirifides, M., Simpson, K., Lin, R., Waterhouse, B., 2001. Topographic organization and neurochemical identity of dorsal raphe neurons that project to the trigeminal somatosensory pathway in the rat. J. Comp. Neurol. 435, 325–340.
- Klepper, A., Herbert, H., 1991. Distribution and origin of noradrenergic and serotonergic fibers in the cochlear nucleus and inferior colliculus of the rat. Brain Res. 557, 190–201.
- Koyama, Y., Kayama, Y., 1993. Mutual interactions among cholinergic, noradrenergic and serotonergic neurons studied by iontophoresis of these transmitters in rat brainstem nuclei. Neuroscience 55, 1117–1126.
- Li, H., Mizuno, N., 1997. Single neurons in the spinal trigeminal and dorsal column nuclei project to both the cochlear nucleus and the

inferior colliculus by way of axon collaterals: a fluorescent retrograde double-labeling study in the rat. Neurosci. Res. 29, 135–142.

- Li, L., Priebe, R., Yeomans, J., 1998a. Prepulse inhibition of acoustic or trigeminal startle of rats by unilateral electrical stimulation of the inferior colliculus. Behav. Neurosci. 112, 1187–1198.
- Li, L., Korngut, L., Frost, B., Beninger, R., 1998b. Prepulse inhibition following lesions of the inferior colliculus: prepulse intensity functions. Physiol. Behav. 65, 133–139.
- Li, Y., Kaneko, T., Mizuno, N., 2001. Collateral projections of nucleus raphe dorsalis neurones to the caudate-putamen and region around the nucleus raphe magnus and nucleus reticularis gigantocellularis pars alpha in the rat. Neurosci. Lett. 299, 33–36.
- Lopez-Garcia, J., 1998. Serotonergic modulation of the responses to excitatory amino acids of rat dorsal horn neurons in vitro: implications for somatosensory transmission. Eur. J. Neurosci. 10, 1341–1349.
- Luethke, L., Krubitzer, L., Kaas, J., 1989. Connections of primary auditory cortex in the new world monkey *Saguinus*. J. Comp. Neurol. 285, 487–513.
- Magoul, R., Oblin, A., Calas, A., Araneda, S., 1988. Serotonergic projections to the spinal cord but not those to the olfactory bulb also contain substance P. A combined immunocytochemical and autoradiographic study following retrograde axonal transport of [³H]serotonin labeled products. Neuroscience 26, 959–969.
- Marsh, R.A., Grose, C.D., Wenstrup, J.J., Fuzessery, Z.M., 1999. A novel projection from the basolateral nucleus of the amygdala to the inferior colliculus in bats. Soc. Neurosci. Abstr. 25, 1417.
- Mooney, R., Huang, X., Shi, M., Bennett-Clarke, C., Rhoades, R., 1996. Serotonin modulates retinotectal and corticotectal convergence in the superior colliculus. Prog. Brain Res. 112, 57–69.
- Moore, R., Bloom, F., 1979. Central catecholamine neuron systems: anatomy and physiology of the norepinephrine and epinephrine systems. Annu. Rev. Neurosci. 2, 113–168.
- Moore, R., Halaris, A., Jones, B., 1978. Serotonin neurons of the midbrain raphe: ascending projections. J. Comp. Neurol. 180, 417–438.
- Morest, D., Oliver, D., 1984. The neuronal architecture of the inferior colliculus in the cat: defining the functional anatomy of the auditory midbrain. J. Comp. Neurol. 222, 209–236.
- Moriizumi, T., Hattori, T., 1991. Pallidotectal projection to the inferior colliculus of the rat. Exp. Brain Res. 87, 223–226.
- Moriizumi, T., Leduc-Cross, B., Wu, J., Hattori, T., 1992. Separate neuronal populations of the rat substantia nigra pars lateralis with distinct projection sites and transmitter phenotypes. Neuroscience 46, 711–720.
- Nakaya, Y., Kaneko, T., Shigemoto, R., Nakanishi, S., Mizuno, N., 1994. Immunohistochemical localization of substance P receptor in the central nervous system of the adult rat. J. Comp. Neurol. 347, 249–274.
- Olazábal, U., Moore, J., 1989. Nigrotectal projection to the inferior colliculus: horseradish peroxidase transport and tyrosine hydroxylase immunohistochemical studies in rats, cats, and bats. J. Comp. Neurol. 282, 98–118.
- Oliver, D., 1987. Projections to the inferior colliculus from the anteroventral cochlear nucleus in the cat: possible substrates for binaural interaction. J. Comp. Neurol. 264, 24–46.
- Oliver, D.L., Huerta, M.F., 1992. Inferior and superior colliculi. In: Webster, D.B., Popper, A.N. (Eds.), The Mammalian Auditory Pathway: Neuroanatomy. Springer-Verlag, New York, pp. 168– 222.
- Oliver, D., Morest, D., 1984. The central nucleus of the inferior colliculus in the cat. J. Comp. Neurol. 222, 237–264.
- Paloff, A., Usunoff, K., 2000. Tyrosine hydroxylase-like immunoreactive synaptic boutons in the inferior colliculus of the cat. Ann. Anat. 182, 423–426.

- Paloff, A., Usunoff, K., Hinova-Palova, D., Ivanov, D., 1985. Retinal innervation of the inferior colliculus in adult cats: electron microscopic observations. Neurosci. Lett. 54, 339–344.
- Park, T., Pollak, G., 1993. GABA shapes sensitivity to interaural intensity disparities in the mustache bat's inferior colliculus: implications for encoding sound location. J. Neurosci. 13, 2050–2067.
- Park, T.J., Grothe, B., Pollak, G.D., Schuller, G., Koch, U., 1996. Neural delays shape selectivity to interaural intensity differences in the lateral superior olive. J. Neurosci. 16, 6554–6566.
- Petersen, R., Diamond, M., 2000. Spatial-temporal distribution of whisker-evoked activity in rat somatosensory cortex and the coding of stimulus location. J. Neurosci. 20, 6135–6143.
- Petrov, T., Krukoff, T., Jhamandas, J., 1992. The hypothalamic paraventricular and lateral parabrachial nuclei receive collaterals from raphe nucleus neurons: a combined double retrograde and immunocytochemical study. J. Comp. Neurol. 318, 18–26.
- Pompeiano, M., Palacios, J., Mengod, G., 1992. Distribution and cellular localization of mRNA coding for 5-HT1A receptor in the rat brain: correlation with receptor binding. J. Neurosci. 12, 440–453.
- Rasmussen, K., Strecker, R.E., Jacobs, B.L., 1986. Single unit responses of noradrenergic, serotonergic, and dopaminergic neurons in freely moving cats to simple sensory stimuli. Brain Res. 369, 336–340.
- Robards, M., 1979. Somatic neurons in the brainstem and neocortex projecting to the external nucleus of the inferior colliculus: an anatomical study in the opossum. J. Comp. Neurol. 184, 547–565.
- Rogawski, M.A., Aghajanian, G.K., 1980. Norepinephrine and serotonin: Opposite effects on the activity of lateral geniculate neurons evoked by optic pathway stimulation. Exp. Neurol. 69, 678–694.
- Sato, A., Ohtsuka, K., 1996. Projection from the accommodationrelated area in the superior colliculus of the cat. J. Comp. Neurol. 367, 465–476.
- Shammah-Lagnado, S., Alheid, G., Heimer, L., 1996. Efferent connections of the caudal part of the globus pallidus in the rat. J. Comp. Neurol. 376, 489–507.
- Shinonaga, Y., Takada, M., Ogawa-Meguro, R., Ikai, Y., Mizuno, N., 1992. Direct projections from the globus pallidus to the midbrain and pons in the cat. Neurosci. Lett. 135, 179–183.
- Shipley, C., Carterette, E., Buchwald, J., 1991. The effects of articulation on the acoustical structure of feline vocalizations. J. Acoust. Soc. Am. 89, 902–909.
- Shneiderman, A., Oliver, D., Henkel, C., 1988. Connections of the dorsal nucleus of the lateral lemniscus: an inhibitory parallel pathway in the ascending auditory system? J. Comp. Neurol. 276, 188– 208.
- Sillar, K., Reith, C., McDearmid, J., 1998. Development and aminergic neuromodulation of a spinal locomotor network controlling swimming in Xenopus larvae. Ann. N.Y. Acad. Sci. 860, 318–332.
- Simmons, J.A., Fenton, M.B., O'Farrell, M.J., 1979. Echolocation and pursuit of prey by bats. Science 203, 16–21.
- Simmons, J.A., Dear, S.P., Ferragamo, M.J., Haresign, T., Fritz, J., 1996. Representation of perceptual dimensions of insect prey during terminal pursuit by echolocating bats. Biol. Bull. 191, 109–121.
- Simmons, J.A., Lavender, W.A., Lavender, B.A., Childs, J.E., Hulebak, K., Rigden, M.R., Sherman, J., Woolman, B., O'Farrell, M.J., 1978. Echolocation by free-tailed bats (*Tadarida*). J. Comp. Physiol. 125, 291–299.
- Stanley, G., Li, F., Dan, Y., 1999. Reconstruction of natural scenes from ensemble responses in the lateral geniculate nucleus. J. Neurosci. 19, 8036–8042.
- Steinbusch, H., 1981. Distribution of serotonin-immunoreactivity in the central nervous system of the rat-cell bodies and terminals. Neuroscience 6, 557–618.

- Tanaka, M., Okamura, H., Yanaihara, N., Tanaka, Y., Ibata, Y., 1993. Differential expression of serotonin and [Met]enkephalin-Arg6-Gly7-Leu8 in neurons of the rat brain stem. Brain Res. Bull. 30, 561–570.
- Thompson, A., Thompson, G., 2001. Serotonin projection patterns to the cochlear nucleus. Brain Res. 907, 195–207.
- Thompson, A.M., Moore, K.R., Thompson, G.C., 1995. Distribution and origin of serotoninergic afferents to guinea pig cochlear nucleus. J. Comp. Neurol. 351, 104–116.
- Thompson, G.C., Thompson, A.M., Garrett, K.M., Britton, B.H., 1994. Serotonin and serotonin receptors in the central auditory system. Otolaryngol. Head Neck Surg. 110, 93–102.
- To, Z., Bonhaus, D., Eglen, R., Jakeman, L., 1995. Characterization and distribution of putative 5-ht7 receptors in guinea-pig brain. Br. J. Pharmacol. 115, 107–116.
- Trulson, M.E., Jacobs, B.L., 1979. Raphe unit activity in freely moving cats: correlation with level of behavioral arousal. Brain Res. 163, 135–150.
- Trulson, M.E., Trulson, V.M., 1982. Differential effects of phasic auditory and visual stimuli on serotonergic neurons in the nucleus raphe dorsalis and nucleus raphe pallidus in freely moving cats. Neurosci. Lett. 32, 137–142.
- Tsunoda, K., Yamane, Y., Nishizaki, M., Tanifuji, M., 2001. Complex objects are represented in macaque inferotemporal cortex by the combination of feature columns. Nat. Neurosci. 4, 832–838.
- Vergé, D., Calas, A., 2000. Serotoninergic neurons and serotonin receptors: gains from cytochemical approaches. J. Chem. Neuroanat. 18, 41–56.
- Waeber, C., Sebben, M., Nieoullon, A., Bockaert, J., Dumuis, A., 1994. Regional distribution and ontogeny of 5-HT4 binding sites in rodent brain. Neuropharmacology 33, 527–541.
- Wallis, D., 1994. 5-HT receptors involved in initiation or modulation of motor patterns: opportunities for drug development. Trends Pharmacol. Sci. 15, 288–292.
- Wang, X., Robertson, D., 1997. Effects of bioamines and peptides on neurones in the ventral nucleus of trapezoid body and rostral periolivary regions of the rat superior olivary complex: an in vitro investigation. Hear. Res. 106, 20–28.
- Waterhouse, B., Moises, H., Woodward, D., 1986. Interaction of serotonin with somatosensory cortical neuronal responses to afferent synaptic inputs and putative neurotransmitters. Brain Res. Bull. 17, 507–518.
- Winer, J., Larue, D., Diehl, J., Hefti, B., 1998. Auditory cortical projections to the cat inferior colliculus. J. Comp. Neurol. 400, 147–174.
- Wright, D., Seroogy, K., Lundgren, K., Davis, B., Jennes, L., 1995. Comparative localization of serotonin1A, 1C, and 2 receptor subtype mRNAs in rat brain. J. Comp. Neurol. 351, 357–373.
- Wynne, B., Robertson, D., 1996. Localization of dopamine-beta-hydroxylase-like immunoreactivity in the superior olivary complex of the rat. Audiol. Neurootol. 1, 54–64.
- Wynne, B., Robertson, D., 1997. Somatostatin and substance P-like immunoreactivity in the auditory brainstem of the adult rat. J. Chem. Neuroanat. 12, 259–266.
- Wynne, B., Harvey, A., Robertson, D., Sirinathsinghji, D., 1995. Neurotransmitter and neuromodulator systems of the rat inferior colliculus and auditory brainstem studied by in situ hybridization. J. Chem. Neuroanat. 9, 289–300.
- Yasui, Y., Kayahara, T., Kuga, Y., Nakano, K., 1990. Direct projections from the globus pallidus to the inferior colliculus in the rat. Neurosci. Lett. 115, 121–125.
- Ye, Y., Kim, D., 2001. Connections between the dorsal raphe nucleus and a hindbrain region consisting of the cochlear nucleus and neighboring structures. Acta Otolaryngol. 121, 284–288.